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Product Sheet

Mouse_FcRn MDCK Cell Line

Catalog number: GM-C34585

Version 3.3.1.250113

Mouse_FcRn MDCK Cell Line is a clonal stable MDCK cell line that constitutively

Description expresses the mouse FcRn(FCGRT) and mouse B2M genes, constructed using lentiviral

technology.

Quantity 3E6 Cells per vial,1 mL

Product Format 1 vial of frozen cells

Shipping Shipped on dry ice

Storage Conditions Liquid nitrogen immediately upon receipt

Target N-Flag-Mouse_FcRn(FCGRT) & Mouse_B2M

Gene ID/Uniprot ID Q61559 & P01887

Host Cell MDCK

Recovery Medium MEM+10% FBS+1% P.S

Growth medium MEM+10% FBS+1% P.S +2.5 μg/mL Puromycin

Note None

Freezing Medium 90% FBS+10% DMSO

Growth properties Adherent

Growth Conditions 37°C, 5% CO₂

Mycoplasma Testing The cell line has been screened to confirm the absence of Mycoplasma species.

Safety considerations Biosafety Level 2

Note

It is recommended to expand the cell culture and store a minimum of 10 vials at an early

passage for potential future use.



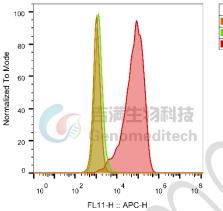
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Materials

Reagent	Manufacturer/Catalogue No.
MEM	Viva Cell/C3050-0500
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Puromycin	Genomeditech/GM-040401
Anti-Flag mIgG1 Antibody	Genomeditech/GM-30726AB

Figures



SampleID	Geometric Mean : FL11-H
MDCK anti-Flag+APC-2nd Ab	1089
MDCK Mouse_FcRn M_IgG+APC-2nd Ab	1380
MDCK Mouse_FcRn anti-Flag+APC-2nd Ab	60599

Figure 1 | Mouse_FcRn MDCK Cell Line (Cat. GM-C34585) was determined by flow cytometry using Anti-Flag mIgG1 Antibody (Cat. GM-30726AB).

Cell Recovery

Recovery Medium: MEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at -70°C. Storage at -70°C will result in loss of viability.

- a) Thaw the vial by gentle agitation in a 37°C water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 3 minutes).
- b) Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- c) Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately 176 x g for 5 minutes. Discard supernatant.
- d) Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.



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e) Incubate the culture at 37°C in a suitable incubator. A 5% CO₂ in air atmosphere is recommended if using the medium described on this product sheet.

Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- a) Centrifuge at 176 x g for 3 minutes to collect cells.
- b) Resuspend the cells in pre-cooled freezing medium and adjust the cell density to 2E6 cells/mL.
- c) Aliquot 1 mL into each vial.
- d) Place the vial in a controlled-rate freezing container and store at -80°C for at least 1 day, then transfer to liquid nitrogen as soon as possible.

Cell passage

Growth medium: MEM+10% FBS+1% P.S +2.5 µg/mL Puromycin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- a) Remove and discard culture medium.
- b) Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- c) Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to the dish to rinse and then aspirate. Repeat this step once. Then, add 2.0 mL of 0.25% (w/v) Trypsin-EDTA solution and observe the cells under an inverted microscope until the cell layer is dispersed (usually within 15 to 20 minutes at 37°C).
- d) Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach.

 Cells that are difficult to detach may be placed at 37°C to facilitate dispersal.
- e) Add 3.0 mL of growth medium to mix well and aspirate cells by gently pipetting.
- f) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- g) Incubate cultures at 37°C.

Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:4 is recommended

Medium Renewal: Every 2 to 3 days

Notes

a) After the stabilization of the cell condition, there will be fewer dead cells post-passage, the cell growth rate will tend to stabilize, cell morphology will become uniform, and the cells will appear robust.

Sequence

Flag-FcRn(FCGRT) Q61559

MGMPLPWALSLLLVLLPQTWGYKDDDDKGSGSETRPPLMYHLTAVSNPSTGLPSFWATGWLGPQQYLTYN SLRQEADPCGAWMWENQVSWYWEKETTDLKSKEQLFLEALKTLEKILNGTYTLQGLLGCELASDNSSVPTA VFALNGEEFMKFNPRIGNWTGEWPETEIVANLWMKQPDAARKESEFLLNSCPERLLGHLERGRRNLEWKEP



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PSMRLKARPGNSGSSVLTCAAFSFYPPELKFRFLRNGLASGSGNCSTGPNGDGSFHAWSLLEVKRGDEHHYQ CQVEHEGLAQPLTVDLDSSARSSVPVVGIVLGLLLVVVAIAGGVLLWGRMRSGLPAPWLSLSGDDSGDLLPG GNLPPEAEPQGANAFPATS

B2M P01887

MARSVTLVFLVLVSLTGLYAIQKTPQIQVYSRHPPENGKPNILNCYVTQFHPPHIEIQMLKNGKKIPKVEMSD MSFSKDWSFYILAHTEFTPTETDTYACRVKHASMAEPKTVYWDRDM

Related Products

FcγR			
Cynomolgus_FcRn MDCK Cell Line	H_FCGR1A(CD64) CHO-K1 Cell Line		
H_FCGR1A(CD64) HEK-293 Cell Line	H_FCGR2A(CD32A) CHO-K1 Cell Line		
H_FCGR2B(CD32B) CHO-K1 Cell Line	H_FCGR3A(CD16a) 158F CHO-K1 Cell Line		
H_FCGR3A(CD16a) 158V CHO-K1 Cell Line	H_FCGR3B(CD16b) CHO-K1 Cell Line		
H_FcRn CHO-K1 Cell Line	H_FcRn MDCK Cell Line		
Anti-FcRn hIgG4 Reference Antibody(Rozabio)	Anti-H_FcRn IgG4 Antibody(Rozanolixizumab)		
Anti-Mouse CD1632 mIgG2b Antibody(2.4G2)	All		
ADCCP			
ADCC FcγRIIIa(158F) Jurkat Effector Cell Line	ADCC FcyRIIIa(158V) DDX35TM Jurkat Effector Cell Line		
ADCC FcγRIIIa(158V) Jurkat Effector Cell Line	ADCC M_FcγRIV Jurkat Effector Cell Line		
ADCP FcγRIIa DDX35TM Jurkat Effector Cell Line	ADCP FcyRIIa Jurkat Effector Cell Line		
ADCP FcyRIIa R131 Jurkat Effector Cell Line	ADCP FcyRIIb Jurkat Effector Cell Line		

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